

Communication to the Editor

ENZYMIC SYNTHESIS OF VALIENAMINE
GLUCOSIDES AND THEIR
ANTIBIOTIC ACTIVITY

Sir:

Although glycosidases have long been known to catalyse the stereospecific formation of glycosidic bonds by reversed hydrolysis or transglycosidation, the predominant formation of (1~6) linkages and the difficulties in the isolation of products have prevented their wider use for synthesis of biologically active carbohydrates. However, the regioselectivity of glycosidase-catalysed formation of oligosaccharides can be manipulated in certain situations by using the appropriate glycosidases and acceptors, and the isolation of products simplified in many reports^{1~3}. We also have reported on microbial glycosidation to form validamycin α - and β -D-glucoside analogs with the yeast, *Rhodotorula lactosa* IFO 1424^{4,5}. Thus, we attempted to prepare α - and

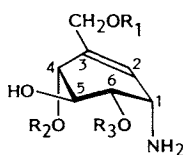
β -D-glucosides of valienamine with glucosidases.

Valienamine (Fig. 1), a pseudo-aminosugar produced by the microbial degradation of validamycins by several soil bacteria^{6,7}, is a competitive α -glucosidase inhibitor, active against yeast- α -glucosidase, invertase and *Rhizopus*- α -glucoamylase, and it shows some antibiotic activity against *Bacillus* sp.⁸

This report deals with the enzymic preparation of α - and β -D-glucoside analogs of valienamine and their antibiotic activity.

For the preparation of partially purified α - and β -glucosidases of *R. lactosa*, cell suspension (400 g) in 20 mM sodium phosphate buffer (pH 6.0) was ruptured with a sonic oscillator and the cell debris were removed by centrifugation at 13,000 $\times g$ for 10 minutes. Ammonium sulfate was added to the supernatant and the precipitate at 0.9 saturation was dissolved in 5 mM phosphate buffer. The solution was put on a DEAE-cellulose column (2.6 \times 95 cm)

Fig. 1. Structure of α - and β -D-glucosides of valienamine.



	R ₁	R ₂	R ₃
Valienamine	H	H	H
VE- α -1	H	α -Glc	H
VE- α -2	H	α -Isomal	H
VE- α -3	α -Glc	H	H
VE- α -4	α -Isomal	H	H
VE- β -1	β -Glc	H	H
VE- β -2	H	H	β -Glc
VE- β -3	H	β -Glc	H

Glc: D-Glucosyl, Isomal: D-isomaltosyl.

Table 1. Selected physico-chemical properties of α - and β -D-glucosides of valienamine.

Compound	$[\alpha]_D^{25}$ (H ₂ O)	TLC Rf ^a	GLC Rt ^b (minutes)	¹ H NMR, in D ₂ O ^c anomeric proton (δ , J = Hz)
VE- α -1	+126.4°	0.55	2.6	5.41 (3.9)
VE- α -2	+160.6°	0.33	27.2	5.43 (3.7), 4.96 (3.7)
VE- α -3	+144.4°	0.48	3.7	4.91 (3.7)
VE- α -4	+137.6°	0.27	33.9	4.94 (3.8), 4.92 (3.7)
VE- β -1	+31.8°	0.46	3.7	4.9 (8.1)
VE- β -2	+30.2°	0.57	2.5	4.64 (7.7)
VE- β -3	+28.4°	0.57	2.7	4.64 (8.3)
Valienamine	+81.6°	0.68		

^a Solvent: CHCl₃ - MeOH - 29% NH₄OH (1:3:2), Silica gel G.

^b 7% OV-17, 2 m, 280°C TMS-derivatives.

^c At 400 MHz with TMS standard.

Table 2. ^{13}C NMR data of α - and β -D-glucosides of valienamine.

Carbon	VE	α -Glucosides				β -Glucosides		
		VE- α -1	VE- α -2	VE- α -3	VE- α -4	VE- β -1	VE- β -2	VE- β -3
C-1	51.7	51.0	51.0	51.8	51.8	51.8	51.1	51.2
C-2	127.2	128.0	128.7	129.6	128.8	129.2	126.0	128.5
C-3	142.0	140.6	140.2	139.5	140.0	140.0	142.6	140.5
C-4	74.7	<u>79.5</u>	<u>79.8</u>	74.1	73.8	74.3	73.7	<u>84.7</u>
C-5	74.9	73.5	74.2	74.8	74.7	74.7	74.0	73.5
C-6	72.9	72.2	73.6	72.4	72.2	72.3	<u>82.7</u>	72.2
C-7	64.1	64.5	64.6	<u>68.9</u>	<u>68.8</u>	<u>72.6</u>	64.1	64.3
C-1'		100.6	100.8	99.1	99.1	104.9	106.2	106.3
C-2'		74.1	74.2	73.9	74.2	75.9	76.0	76.1
C-3'		75.7	75.8	75.8	76.0	78.4	78.3	78.4
C-4'		72.1	72.1	72.3	72.2	72.4	72.1	72.1
C-5'		75.4	72.4	74.6	73.0	78.6	78.5	78.7
C-6'		63.2	<u>68.5</u>	63.2	<u>68.2</u>	63.4	63.2	63.4
C-1''			100.6		100.6			
C-2''			74.0		74.0			
C-3''			75.9		75.8			
C-4''			72.2		72.0			
C-5''			74.5		74.6			
C-6''			63.2		63.2			

Chemical shifts are in ppm downfield of DSS. ^{13}C NMR spectra were taken in D_2O on a Jeol JNM-GX 400 spectrometer.

equilibrated with the buffer. The column was washed with 10 mM buffer and eluted with the buffer containing 50 mM NaCl. The α -glucosidase fraction (72.6 U/maltose, 0.12 U/mg · protein) and β -glucosidase fraction (63.5 U/cellobiose, 0.22 U/mg · protein) were obtained.

α -(or β -)Glucosidation reaction of valienamine was accomplished as follows. Reaction mixture (500 ml) containing valienamine (2.5 g), maltose (100 g, or cellobiose 50 g) and the α -glucosidase fraction (50 U, or the β -glucosidase fraction) in 40 mM acetate buffer (pH 5.0) was incubated at 27°C for 72 hours. The glycosidation process was followed by TLC (silica gel, CHCl_3 - MeOH - 29% NH_4OH , 1:3:2) and gas-liquid chromatography (GLC, trimethylsilyl derivatives, 7% OV-17 on Chromosorb AW, 280°C⁹). The products were isolated by column chromatography on Dowex 50WX8 (H form, 1.6 × 60 cm), eluted with 0.5 N NH_4OH and then on Dowex 1X2 (OH form, 1.0 × 70 cm) and eluted with water. The reaction of α -glucosidation gave four components, VE- α -1 (301 mg), VE- α -2 (90 mg), VE- α -3 (32 mg) and VE- α -4 (22 mg) as amorphous materials which are distinguishable from each other and homogeneous by the TLC and GLC. β -Glucosidation gave three components, VE- β -1 (180 mg), VE- β -2 (138 mg) and VE- β -3 (60 mg). Selected physico-chemical properties and NMR

Table 3. Antibacterial activity of valienamine glucosides in agar diffusion assay with nutrient broth.

Compound	Diameter of inhibition zones (10 mg/ml) ^a		
	<i>Bacillus subtilis</i>	<i>B. cereus</i>	<i>B. megaterium</i>
VE- α -1	25 mm	21 mm	23 mm
VE- α -2	20	19	20
VE- α -3	26	19	24
VE- α -4	13	+	12
VE- β -1	0	0	0
VE- β -2	0	0	0
VE- β -3	0	0	0
Valienamine	(26) ^b	(21) ^b	(24) ^b

^a Nutrient agar (Nissui), 24 hours incubation at 37°C, compound 500 μg /disc (diameter 8 mm).

^b Indicates inhibitory zones were not clear.

chemical shifts of the valienamine glucosides are listed in Tables 1 and 2. Complete signal assignment of ^{13}C NMR spectra was almost made by comparing each spectrum with corresponding α -glucotrioses and α -glucobioses^{10,11}. Their shift patterns were consistent with the known effects of glucosidation (underlined in Table 2), including β - and γ -effects of the olefinic carbons at the allylic position¹². The ^{13}C resonance shifts produced by changes of pD were useful for the assignment. Deuteration of

amino groups caused large upfield shifts of the signals which are β - to the amino groups¹³⁾. For example, the chemical shifts of C-2 and C-6 in VE- β -2 were shifted upfield by 7.9 and 4.6 ppm, respectively by protonation. Further confirmation of spectral assignment was achieved from ¹³C-¹H COSY experiments. The structures are depicted in Fig. 1.

As shown in Table 3, the α -D-glucosides of valienamine have some antibacterial activity against *Bacillus* sp. on nutrient agar medium by the paper-disc method (concentration 10 mg/ml, 500 μ g/disc) as well as valienamine and showed clear inhibition zones, while valienamine showed hazy zones, incomplete inhibition. However, the β -D-glucosides showed no inhibitory zone.

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